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Lesson 13: Functional Genomics II: Functional Genomics Databases and Overrepresentation Analysis and Clustering of Microarray Data

In this class we are going to learn the basics of using web-accessible function and pathway databases, and how to find the functions and pathways associated with a set of differentially expressed genes.

Theory

1. Pevsner on functional databases:

A. OMIM p. 659-661. B. GO 243-246. C. KEGG 258-263.

2. Ontools. Read sections on Onto-Express and Pathway-Express.

http://vortex.cs.wayne.edu/projects.htm

3. Pevsner on clustering or microarray data: p. 203-214.

Theory of Hierarchical Clustering (Unsupervised Learning)

 d_{xy} = distance in expression patterns between genes x and y.

$$d_{xy} = 1 - r_{xy} = 1 - \frac{1}{N} \sum_{i=1,N} \left(\frac{x_i}{\sqrt{\sum_{i=1,N} x_i^2}} \frac{y_i}{\sqrt{\sum_{i=1,N} y_i^2}} \right)$$

 r_{xy} = modified Pearson correlation between expression of genes x and y with zero mean. N= Number of samples.

Then a tree is inferred based on distances.

Summary of Commands:

Note: In this document different fonts have different meanings:

Times is used to explain commands.

Courier is used to indicate commands and command options.

Courier italics are used to indicate command parameters, for example, filenames.

Courier bold is used to indicate commands that are not displayed.

Courier bold italics are used to indicate computer-generated output.

Helvetica is used to indicate menu items.

http://www.ncbi.nlm.nih.gov/entrez	Accesses the NCBI ENTREZ web page
Gene	Diverse information on genes including
	molecular interactions.
OMIM	Descriptive genotype-phenotype
	relationships in humans.
OMIA	Descriptive genotype-phenotype
	relationships in organisms other than
	humans.
http://www.geneontology.org/	Gene Ontology Gives Biological Process,
	Molecular function, Cellular Component for
	proteins/
Gene symbol/name, exact match	To search for a gene/protein
	This will give you all of the terms associated
	with a protein.
Terms	To search for a term. Following the
	links associated with a term will give
	you a definition of the term and the names
	of all of the proteins associated with that
	term.
http://bond.unleashedinformatics.com/Action?	
BOND: Biomolecular Object Network Database.	
	A database that gives interaction
	partners between biological molecules.
Search bind using a field specific query	Enables user to specify the kind of search.
Field: Interaction description contains at	least one of the following words
	Generates a list of interactions in which a
	specified molecule plays a role.
On the list page clicking on an	
interaction identifier	gives a description of the interaction in
	gives a description of the interaction in
	question.

On the interaction page, clicking on: expand all

http://www.genome.jp/kegg/

Click on KEGG Table of Contents KEGG PATHWAY KEGG GENES SEARCH KEGG for *subject* GENES for *genename*

PATHWAYS for *pathway*

SEARCH organismcode for genename

Organism GENES for *Genename* PATHWAYS for *Pathway* gives a more detailed picture of the interaction in question.

KEGG: Kyoto Enyclopedia of Genes and Genomes.

Encyclopedia of Pathways. Encyclopedia of Genes.

Searches Gene database for name of gene. Searches Gene database for name of gene. Searches Pathway database for name of pathway.

Searches for name of gene in specific organism.

Searches for organism code.

Searches Gene database for name of gene. Searches Pathway database for name of pathway.

Analysis with Onto-tools

- A. Preprocessing for ontools:
 - 1. Move output files to a directory that you can work in.
 - 2. Open output files with Microsoft Excel. Autoformat column length and save file as Excel Workbook with new name.
 - 3. Copy all genes with B > 0 to new Excel Workbook and save as Workbook with new name.
 - 4. Delete all columns but the Affymetrix probeset identifier. Delete the header row. save as text file.
- B. Analysis with Onto-Express and Pathway-Express.

 <u>http://vortex.cs.wayne.edu/ontoexpress/</u>

 Login:
 Login to web-site.

 Onto-express
 Select Onto-Express for finding

overrepresented Gene Ontology Categories.

Fill out the ontoexpress input window as below (settings for experiment- the parameters will of course vary with other files, organisms, and chip).

Save Directory: uments\sequencecourse\lesson13 Save: <	Save Results	
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Onto-express output files: bioprocess

molecular

cellular

chromosome

To process output files with Excel: Open Files as type *textfiles* Delimited Tab Semi-colon Format->Column->Autofit Overrepresented biological process categories. Overrepresented molecular function categories. Overrepresented cellular location categories. Assignment of genes to chromosomes.

Opens file from inside Excel. Lists text files in the menu. Reads file as entried separated by delimiters. Recognizes tabs as delimiters. Recognizes semi-colons as delimiters. Fits selected columns.

Data->Sort	Sorts by the selected column.
Expand Selection	Sorts rest of row according to the selected
	column.
File->Save As	Save as file with name typed in box.
Save As type <i>Microsoft</i>	Excel Workbook
	Save as file in Excel Format.

1. Open each of these files with Excel (check delimited and tab and semicolon). Autofit selection for column, sort by corrected p-value, and save as a Workbook.

Save Results	
Save Directory:	uments\sequencecourse\lesson13 Browse
Save:	
	Rological Process 🔽 Cellular Component
<u>e</u> c	ni omosome Location 🕑 Molecular Function
✓ Save Details	✓ Print Headers
	🗹 Accession Number 🔽 Locus ID
	🗹 Cluster ID 🗾 🗹 Official Gene Symbol
✓ Save Tree Vie	w
	Save Cancel
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The above box will output the same 4 files as before only with the genes included. In addition, it will output, "treeview", a file with all of the gene ontology categories detected and "treeview_input", a file with all of the gene-ontology categories detected along with input genes from the list. The above files cannot be sorted by corrected p-values without getting mixed up, because there is more than one row per Gene Ontology category.

Pathway-Express

Select Pathway-Express for finding overrepresented KEGG pathwayCategories. Use list of pathways with log fold changes for input.

SeathwayExpress	- Input								
-Run a demo dataset									
Select a demo dataset Demo Info									
	Run <u>D</u> emo								
Analyze your own dataset									
Input File :	iencecourse\lesson13\10p-vbgt0.txt	Input File							
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Organism :	Homo sapiens 🔹								
Input Type : 🤇	Gene Symbol OLocusLink								
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Advanced options — Advanced optior	IS								

Advanced options Corrections = fdr

	Corrections = fdr	Corrects pathways for false discoveries with false discovery rate.					
Pathw	ay express output-windows and options	-					
Bar G	iraph	Display of overrepresented pathways					
Pathv	vay Details	Shows list of pathways in tables.					
	(Right mouse button over pathway link)	Lists pathway options.					
	Show Pathway Genes	Changes pathway in Pathway Gene Details.					
	Show Pathway Details	Shows KEGG diagram of pathways with genes on list indicated.					
	Save Table	Saves Pathway Details file.					
Pathv	vay Gene Details	Shows list of genes in selected pathway and indicates whether or not they are part of the input list.					
	(Right mouse button over genelink)	Lists gene options options, which are analogous to Pathway Details options.					
Input	Details	Shows list of genes in input lists and indicates whether or not they are part of a pathway. Mouse options analogous to other windows.					

Hierarchical clustering with Cluster 3.0 and JavaTreeview.

1. Start R and go to the estrogen working directory.

R RGui	🔳 🖻 🗙
File Edit View Misc Packages Windows Help	
Ŗ R Console	
R version 2.6.1 (2007-11-26) Copyright (C) 2007 The R Foundation for Statistical Computing ISBN 3-900051-07-0	A.
R is free softwar Choose directory NTY. You are welcome t Type 'license()' Change working directory to: 115.	
Natural languag R is a collaborat Type 'contributor OK Canvel	
Type 'demo()' for some demos, 'help()' for on-lime help, or 'help,start()' for an HTML browser interface to help. Type 'q()' to quit R.	
<pre>> local((pkg <- select.list(sort(.packages(all.available = TRUE))) + if(nchar(pkg)) library(pkg, character.only=TRUE))) > </pre>	
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2. Load the Affy Program:



3. In R window:

```
estrogenEset<-ReadAffy() Loads Estrogen cell files into Expression
object.
estrogenmas5 <-mas5(estrogenEset) Normalizes cel files by the MAS5
algorithm.
write.exprs(estrogenmas5,"estrogenmas5.txt")
Saves normalized arrays as text file.
```

- 4. Cut and paste file to directory which you are working in for lab.
- 5. Copy file with differential expression values from last week to file called id_gene.
 - A, Open it with excel.
 - B. Delete all columns other than Gene-and-symbol.
 - C. Sort by ID.
 - D. Rename Symbol Column "Gene".

The spreadsheet should look like this:



6. Open Estrogenmas5 within Excel.

A. Make 2 new columns.

B. Paste in the contents of ID gene so that the id column should be aligned The file should look like this:

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4	1001_at	1001_at	TIE1	15.89701	53.2984	22.50213	132.8748	39.70776	18.18771	155.9373	105.1721			
5	1002_f_at	1002_f_at	DID1	105,0000	20.47404	32.03002	90.04274	22.05010	39.21241	47.00420	41.11254			
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16	1012 at	1012 at	PCAF	70.72225	62.1532	22.92618	91.96284	83.4887	77.49091	40.00592	118.0775			
17	1013 at	1013 at	SMAD5	113.7053	116.7283	48.62628	39.85557	114.7546	79.01453	47.96287	6.717195			
18	1014 at	1014 at	POLG	1084.494	765.7347	1548.439	1162.647	1080.62	1143.464	1312.972	1234.014			
19	1015_s_at	1015_s_a	t LIMK1	46.45188	24.98275	130.6936	131.872	52.44243	146.2038	231.3765	350.6037			
20	1016_s_at	1016_s_a	t IL13RA2	52.10842	33.36456	43.2515	66.7575	13.58892	46.67975	20.00002	110.7657			
21	1017_at	1017_at	MSH6	93.96083	81.7148	126.1752	86.70582	89.14314	88.27552	182.2691	120.9935			
22	1018_at	1018_at	WNT10B	87.93711	76.41729	57.79347	250.9544	61.85392	88.98151	51.78122	122.5474			
23	1019_g_at	1019_g_a	t WNT10B	217.5104	230.4868	367.7689	514.2963	243.2797	287.2168	393.8499	290.5965			
24	102_at	102_at	HIPK3	38.99282	39.24108	52.02848	167.2717	61.05234	56.144	133.0828	67.93559			
25	1020_s_at	1020_s_a	t CIB1	2184.607	2755.012	2120.54	1702.346	3209.82	3336.758	2896.835	2499.176			
26	1021_at	1021_at	IFNG	39.1303	25.57162	62.59795	135.6775	87.51481	46.20445	84.75281	124.9427			
27	1022_f_at	1022_f_at	IFNA14	33.17248	52.70055	5.195407	105.0966	45.18682	21.55817	23.65856	7.845053			
28	1023 at	1023 at	EPO	37.10382	36.06208	40.79993	49.4669	23.78648	20.15079	30.35107	54.32572			

7. Delete the 2 id columns so that the spreadsheet looks like this:

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10	DDR1	5261.891	6496.665	5145.718	5207.922	8383.247	7866.76	7706.648	6125.094						
11	EIF2AK2	4198.788	922.6769	3042.358	5653.327	1634.609	5124.613	4401.711	6235.49						
12	HINT1	4598.059	4052.111	6772.822	4628,419	2966.029	2789.746	2196.602	1670.843						
13	DYRK4	17.22251	15.34156	24.00909	17.15284	23.24743	15.41839	19.59627	23.04363						
14	MAPK11	17.46486	61.21554	20,76101	44.85587	17.36519	39.68644	88.85223	76.91898						
15	YWHAE	907.1976	767.2434	2211.709	1484.353	732.8722	856.4684	1032.797	721.4463						
16	PCAF	70.72225	62.1532	22.92618	91.96284	83.4887	77.49091	40.00592	118.0775						
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18	POLG	1084.494	765.7347	1548.439	1162.647	1080.62	1143.464	1312.972	1234.014						
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23	WNT10B	217.5104	230.4868	367.7689	514.2963	243.2797	287.2168	393.8499	290.5965						
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25	CIB1	2184.607	2755.012	2120.54	1702.346	3209.82	3336.758	2896.835	2499.176						
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8. Save file as text file.called "estrogenmas5gene"

9. Open Cluster 3.0 on the PC and load mas5gene



10. Click adjust data. Check

- A. Log Transform
- B. Center Genes
- C. Center Arrays



D. Click Apply

Click filter data

11. In filter data Check

- A. SD (Gene Vector) 2.0
- B. At least one observation with $abs(val) \ge 2.0$
- C. MaxVal-MinVal>=2.0

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D. Click Apply filter. The GUI should say "70 passed out of 12625"

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. "	V SD (Gene Vector) 20	Search for:
1	At least 1 observations with abs[Val] >= 2.0	Example: "Print more than one copy"
	₩ MaxVal - MinVal >= 2.0	Open Romartine Alement
Page 8	Apply Filter	
	70 passed out of 12625	
	Accept Filter	
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E. Click Accept filter

12, Click on Hierarchical tab. Check

- A. Cluster Genes.
- B. Cluster Arrays

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Then click "Average linkage"

Files entitled:

A.	estrogenmas_gene.atr	(ATR)
B.	estrogenmas_gene.	(CDT)
C.	estrogenmas_gene.	(ATR)

Should appear in your directory. You should now be ready to display the cluster diagram and heatmap.

13. Java Treeview

- A. Open Java Treeview.B. Select Open-> File
- C. Scroll to directory containing CDT file
- D. Open CDT file

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14. You should get a heat map that looks like this.

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15. You can select and magnify clusters by clicking on nodes:



16. The resulting heat map has good expansion for exploring clusters.



Lab

1. Characterize the function, interactions and pathways of the c-src (or the protein of your choice) using web-accessible databases. Save the necessary information.

2. What is the phenotype associated with the BRCA1 185DELAG mutation in humans?3. From the list of differentially expressed genes obtained in the 10 hour estrogen experiment explained in part I, generate the following lists in Excel Workbook format:

- A. The Biological Function Gene-ontology values overrepresented in the Estrogen experiment sorted by corrected p-values.
- B. The Biological Function Gene-ontology values overrepresented in the Estrogen experiment with gene symbols and other identifiers (unsorted).
- C. The KEGG pathways overrepresented in the Estrogen experiment.

4. Produce a heatmap representing the gene expression patterns of the estrogen dataset from the last lab clustered by both gene and array.

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