Document Change History

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<th>Version Number</th>
<th>Date</th>
<th>Contributor</th>
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<td>V1.0 draft</td>
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<td>Kenneth C. Smith</td>
<td>This supplement to the geWorkbench manual describes several new features, including caGrid Services, caScript, and a caArray query interface.</td>
</tr>
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<tr>
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<th>User’s Guide</th>
<th>Program Management</th>
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<tr>
<td>Names of developers</td>
<td>Names of technical writers and reviewers</td>
<td>Names of program managers</td>
</tr>
<tr>
<td>Andrea Califano</td>
<td>Kenneth Smith</td>
<td>Aris Floratos</td>
</tr>
<tr>
<td>Aris Floratos</td>
<td>Eileen Daly</td>
<td>Kenneth Smith</td>
</tr>
<tr>
<td>Matt Hall</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Michael Honig</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Bernd Jagla</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Kiran Keshav</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Manjunath Kustagi</td>
<td></td>
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</tr>
<tr>
<td>John Watkinson</td>
<td></td>
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<tr>
<td>Xiaoqing Zhang</td>
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¹ All contributors are currently members of the Joint Centers for Systems Biology, Columbia University, New York, NY.
# Contacts and Support

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## LISTSERV Facilities Pertinent to software teams

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Chapter 1  Introduction to the Manual

This manual is intended for users of geWorkbench who wish to experiment with newly added advanced features. It is directed at the bench scientist and bioinformatician. This manual does not provide installation instructions for geWorkbench nor for grid services.

Topics in this introductory chapter include:

- Topics covered in this supplement
- Organization of the Guide
- Getting Started with geWorkbench

Topics covered in this supplement

This manual is a supplement to the geWorkbench User Manual v.1.3, dated September 14, 2006. That manual explains the basic principles, design goals, and uses of geWorkbench, a software platform which is centered around the single or joint analysis of gene expression microarray and sequence data.

This supplement adds material covering newly developed features which provide advanced functionality. Some must still be regarded as experimental. As the software matures, final descriptions will be incorporated directly into the User Manual.

The new software modules described in this supplement relate to

1. use of geWorkbench within the context of the caGRID infrastructure. Several analytical routines already supported directly within geWorkbench have been developed as formal caGRID services, with an appropriate service interface present within geWorkbench. They are initially intended to be used in the analysis of microarray data. They are:

   a. Hierarchical Clustering
   b. SOM (Self-Organizing Maps)
   c. ARACNE (a gene network reverse-engineering tool)

2. a query interface for caARRAY which allows searches on available annotation fields

3. use of the caSCRIPT scripting language developed specifically for geWorkbench to automate the running of repetitive or complex tasks.

This manual will provide detailed examples on how to use these new modules.
## Organization of the Guide

<table>
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<th>Chapter Contents</th>
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<tr>
<td>Chapter 2</td>
<td>A general introduction to geWorkbench, and a high-level description of the advanced services covered in this manual.</td>
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<tr>
<td>Chapter 3</td>
<td>Description of the geWorkbench User Interface, including layout, basic file operations, and using Online Help</td>
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<td>Chapter 6</td>
<td>Using the java-like language scripting language caSCRIPT to automate routine or complex operations.</td>
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<td>Chapter 7</td>
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## Getting Started with geWorkbench

To get started with geWorkbench you may refer to the following sections of this manual:

- This introduction provides an overview of the manual structure
- Review Chapter 2 for a brief overview of the software
- Review Chapter 3 to learn about the Graphical User Interface
- Refer to Chapters 4, 5, and 6 a description of how to use the advanced services covered in this supplement.

Please note that this manual is a supplement to the main geWorkbench User Manual. Additional information about running geWorkbench can be found there. Detailed instructions and step-by-step tutorials on how to install and run geWorkbench are available online at [http://www.geworkbench.org/](http://www.geworkbench.org/).
# Document Text Conventions

The following table shows various typefaces to differentiate between regular text and menu commands, keyboard keys, and text that you type. This illustrates how conventions are represented in this guide.

<table>
<thead>
<tr>
<th><strong>Convention</strong></th>
<th><strong>Description</strong></th>
<th><strong>Example</strong></th>
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<tr>
<td><strong>Bold &amp; Capitalized Command</strong></td>
<td>Indicates a Menu command</td>
<td><strong>Admin &gt; Refresh</strong></td>
</tr>
<tr>
<td>Capitalized command &gt;</td>
<td>Indicates Sequential Menu commands</td>
<td></td>
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<tr>
<td>Capitalized command</td>
<td></td>
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</tr>
<tr>
<td>TEXT IN SMALL CAPS</td>
<td>Keyboard key that you press</td>
<td><strong>Press ENTER.</strong></td>
</tr>
<tr>
<td>TEXT IN SMALL CAPS + TEXT IN</td>
<td>Keyboard keys that you press simultaneously</td>
<td><strong>Press SHIFT + CTRL and then</strong></td>
</tr>
<tr>
<td>SMALL CAPS</td>
<td></td>
<td><strong>release both.</strong></td>
</tr>
<tr>
<td>Boldface type</td>
<td>Options that you select in dialog boxes or drop-down menus. Buttons or icons that you click.</td>
<td>In the Open dialog box, select the file and click the Open button.</td>
</tr>
<tr>
<td><strong>Italics</strong></td>
<td>Used to reference other documents, sections, figures, and tables.</td>
<td><strong>caCORE Software Development Kit 1.0 Programmer’s Guide</strong></td>
</tr>
<tr>
<td><strong>Italic boldface type</strong></td>
<td>Text that you type</td>
<td>In the New Subset text box, enter <strong>Proprietary Proteins.</strong></td>
</tr>
<tr>
<td><strong>Courier typestyle</strong></td>
<td>Used for filenames, directory names, commands, file listings, source code examples and anything that would appear in a Java program, such as methods, variables, and classes.</td>
<td><strong>URL_definition ::= url_string</strong></td>
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**Note:** Highlights a concept of particular interest

**Warning!** Highlights information of which you should be particularly aware.

**{}** Curly brackets are used for replaceable items.

Replace **{root directory}** with its proper value such as **c:\cabio**

*Table 1. Document Conventions*
Chapter 2  Overview of the Software

This chapter provides an overview of geWorkbench itself, and a description of the particular advanced services that will be covered in this manual: access to remote analytical services via caGRID, a query interface for caARRAY, and scripting with geWorkbench using caSCRIPT.

Topics in this chapter include:

- Introduction to geWorkbench
- Components of geWorkbench
- Features and Functions of the Software
- Brief Description of the User Interface.

Introduction to geWorkbench

geWorkbench is a framework for bioinformatics data analysis. It provides data management, visualization, analysis and retrieval capabilities. It has been primarily constructed for analysis of data derived from gene expression microarray experiments, and allows pulling in many different resources to this end, including sequence, gene ontology, promoter analysis, and standard analytic techniques such as the t-test, hierarchical clustering, and gene network reverse-engineering.

geWorkbench has a modular, component-based design. New modules can easily be written and added as the need arises. A primary aim is to allow easy integration of different forms of bioinformatic data analysis. Such integration of different software routines removes the common hindrance of needing to reformat data for each different type of analysis undertaken.

Extensive documentation and training material for geWorkbench can be found on its main website at http://www.geworkbench.org/. There are wiki-based tutorials there for almost all components of the application. These tutorials are more applied in nature than the material in the printed manual. The software can be downloaded via links found on the 'Download' section of that site. Those links refer to the actual archival location of the software, which is the GForge site maintained by the NCICB. All official releases of the software can be downloaded from that site.
Components of geWorkbench

geWorkbench: geWorkbench v.1.0 is a Java application which is run on the User’s local Windows, Macintosh or Linux workstation. This main application also serves as a front-end client to a number of external computational and data services. Such services already present in geWorkbench include the ability to run BLAST jobs on NCBI servers, and to retrieve gene, pathway and sequence information from sources such as UC Santa Cruz and the NCICB. This supplement also describes caSCRIPT, which is a Java-like scripting language that can be used to automate tasks within geWorkbench. It is an interpreted language which runs within the geWorkbench client.

caGRID: The goal of caGRID is to provide standardized, reusable services. caGRID uses an enhanced variety of web-services to manage communications between the local client, in this case geWorkbench, and remote services. In implementing a framework that is intended to allow programs to interoperate, it provides a mechanism for all data and parameters passed on the grid to be of known, registered types.

geWorkbench is a consumer of grid services. As part of a caGRID proof-of-concept development project, the geWorkbench team has implemented three remote analytical grid services which have been approved at the caBIG silver-level: Hierarchical Clustering, SOM, and ARACNE. These remote services are accessible via caGRID through geWorkbench.

caArray: geWorkbench has previously supported data retrieval from instances of the caArray microarray gene expression database. The latest version of caArray now supports queries against annotations stored in the database concerning arrays and experiments. geWorkbench has implemented an interface based on the MAGE-OM API allowing such queries to be composed and dispatched.

Features and Functions of the Software

Three new sets of functionality are covered in this supplement to the User Manual. For all other topics please see the geWorkbench User Manual v.1.3. The new developments covered here are:

1. use of geWorkbench within the context of the caGRID infrastructure to access remote analytical services.

2. development of a graphical user interface for MAGE-OM API-based queries of caARRAY, which allows searches on available annotation fields.
3. use of the caSCRIPT scripting language developed specifically for geWorkbench to automate the running of repetitive or complex tasks.

The new grid services covered in this supplement are Hierarchical Clustering, Self-Organizing Maps (SOM) and ARACNE. These are routines which can be used in the analysis of microarray data. These routines already existed in geWorkbench and function in the same manner as they did before. The difference is that now the user is provided with the option of discovering and choosing a caGRID node on which to run the computation. To this end, a **Service** tab has been added to the geWorkbench GUI for each routine.

Hierarchical Clustering is used to group a set of genes or microarrays based on similarities in the data. It can be used for example to find groups of genes with similar expression patterns across different tissue types or treatment conditions. Its output is a single connected graph of genes and/or arrays. A graphical viewer in geWorkbench displays the result and allows its manipulation, e.g. retrieving the list of genes from a particular branch of the graph.

SOM analysis divides a data set into a user-chosen number of discreet sets, again based on similarities in the data. Several parameters can be adjusted to determine how this grouping will be performed. Again, a graphical viewer is provided to inspect the results.

ARACNE can be used to find which gene is influencing the expression of another – it is a network reverse-engineering algorithm. It is based on an information-theory calculation known as mutual information. The result is believed to be more robust than a simple correlation calculation. It generally requires at least 100 array experiments for proper sensitivity.

The new geWorkbench query interface for caArray makes use of annotations stored in the database under the MAGE standard. It allows a query to be constructed using several standard fields, such as species, organ, or microarray platform.

cSCRIPT is a Java-like interpreted scripting language that can be used within geWorkbench. Scripts can be stored and reloaded for later execution. caSCRIPT can execute any geWorkbench analysis in the same way as it would be launched interactively from the geWorkbench GUI.
Brief Description of the User Interface

geWorkbench is a Java application. It is currently written to the Sun Java 1.5 (aka Java 5) specification and requires a Java 1.5 JRE to be installed on the client system in order to run. The user interface provides built-in areas for managing data, sub-setting data, viewing data and results, and finally a region for data analysis. There is a built-in Online Help system which covers most standard functionality which has been released as part of the formal release process. One hallmark of the geWorkbench GUI design is that it only displays those components relevant to the currently selected data type. That is, if a gene sequence object is selected in the data window, only components relevant to manipulating sequences will be displayed.
Chapter 3 The geWorkbench User Interface

This chapter provides a basic introduction to using the geWorkbench Graphical User Interface (GUI) as it relates to the advanced services described in this supplement. A more detailed description can also be found in the geWorkbench User Manual v.1.3 and in the online tutorials (http://www.geworkbench.org/).

Topics covered include:

- Basic Layout
- Online Help.
- Working with Data Files

Basic Layout

The graphical user interface for geWorkbench is divided into four major sections:

1. Data management - Workspace and Projects (upper left)
2. Marker and Array/Phenotype set selection and management (lower left)
3. Visualization tools (upper right)
4. Analytical tools (lower right)
Menu Bar

The GUI provides a menu bar at top with a standard choice of commands. Many commands that are available in the menu bar are also available by right-clicking on data objects.

Data management area (1)

Working with geWorkbench involves creating a project within the top-level workspace. Open data files and the results of data transformation or analysis are stored within a project. A workspace can contain more than one project at a time, allowing data to be organized as desired. A workspace and all the projects and data within it can be saved and later reloaded.

Set selection and management (2)

A key feature of geWorkbench is the ability to work with defined sets of markers or arrays. This allows subsets of data to be analyzed, and allows for passing of selected subsets of data between different components. For example, the t-test can be used to create a list of markers showing a significant difference in expression between two states, and this list can then be used to retrieve relevant sequences or annotations.
Visualization and Analysis tools (3 and 4)

geWorkbench works such that only the visualization and analysis components relevant to the type of dataset currently selected in the Project Folders area (1) are displayed through tabs in their respective areas (3 and 4). Thus choosing a microarray dataset will result in a different set of tabs being displayed as compared with those seen when a nucleotide sequence file is selected. When a new data file is loaded, or an analysis produces a new data set, not only is it added to the Project area (1), but an appropriate viewer in the Visualization area (3) is automatically selected.

Online Help

Figure 3-2 shows the Online Help interface. Online Help is found as a menu item under Help on the top menu bar. Online Help is provided for all geWorkbench modules which have been included in a formal release. They focus on the actual use of particular controls within a given module, e.g. button actions, definition of parameters etc.
Working with Data Files

The basics of opening a data file and the details of working with microarray data sets are covered in the geWorkbench User Manual and in the online tutorials (http://www.geworkbench.org/). To aid in the usability of this manual the background needed to work with files will be quickly reviewed.

Prerequisites

Much of the functionality of geWorkbench currently depends on annotation files supplied
by Affymetrix for their microarray chips. Due to licensing restrictions, these files are not
distributed with geWorkbench as part of formal releases. The examples in this
supplement to the User Manual however do not depend on annotation information. If
you nonetheless would like to work with the full functionality of geWorkbench, the
relevant file for the dataset used in this manual can be downloaded from the
Affymetrix.com support web site. The file is named “HG_U95Av2_annot.csv”.

Workspaces, Projects, and Files

The top level of organization of data in geWorkbench is the Workspace. A Workspace
can contain any number of Projects, which are used to organize data and results.

geWorkbench includes sample data files. In the example below we will open a small
microarray data file, web100.exp. This file is in a custom format used by
geWorkbench, which is termed the Affymetrix File Matrix format. It contains results
from a number of different microarray chips that have been merged into one dataset.

Opening a file in a new Project (see Figure 3-3)

1. Right-click on Workspace and select New Project.
2. Right-click on Project and select Open File(s).
3. By default, the file browser should open in the geWorkbench data directory.
   Select File of Type to be Affymetrix File Matrix.
4. Select the file web100.exp.
5. A box with information about annotation files will appear. Click **Continue**.

6. The file browser will open at the root of the geWorkbench installation directory (Figure 3-4). If the file `HG_U95Av2_annot.csv` is present, just press the **Open** button. If you have downloaded it to another directory, please navigate to that directory and open the file. If you do not have the file, just press **Cancel** and proceed without the annotation file.
Chapter 3 User Interface

Figure 3-4 Opening the annotation file

The opened data file is now shown within the **Project Folders** area at upper left in the GUI. All components relevant to acting on microarray data have now appeared in the interface Figure 3-5.
Figure 3-5 geWorkbench GUI showing all microarray-related modules
Chapter 4 Using caGRID Analytical Services

This chapter describes how geWorkbench can be used to dispatch jobs to remote servers using the caGRID infrastructure. Three services which have been implemented are Hierarchical Clustering, Self Organizing Maps, and ARACNE.

Topics covered in this chapter include:

- Using caGRID-based remote analytical services
- Hierarchical Clustering
- Self-Organizing Maps (SOM)
- ARACNE

Using caGRID-based remote analytical services

Three microarray analysis routines already available within geWorkbench have been deployed as caGRID analytical services: Hierarchical Clustering, Self-Organizing Maps (SOM), and ARACNE. The purpose is to remove large-scale calculations from the user’s desktop machine, instead running them on appropriately scaled server systems. The remote systems could scale to cluster computers or other major hardware platforms as demand necessitates. This could be of particular interest for jobs requiring or benefiting from parallel programming, large memory, or which have long run-times. caGRID provides a standardized way to develop, deploy, and interact with these remote services.

Information about the hierarchical clustering and SOM routines is available in either or both of the geWorkbench User Manual and the tutorials available on http://www.geworkbench.org/. This material will not be repeated in detail in this supplement, as the only new feature is the availability of remote execution over caGRID. The material on ARACNE however is new.

This chapter will primarily focus on invoking the grid-based services. Use of all three routines begins with loading a microarray dataset. An example of doing so has already been provided in Chapter 3, using the supplied data file web100.exp. Such datasets are two-dimensional, in that typically results from several experiments (chips) have been merged into a single data array, with genes on the vertical axis and the individual experiments on the horizontal axis when viewed in spreadsheet format.

Hierarchical Clustering

Hierarchical clustering is a method used to group data based on a measure of similarity. The two-dimensional microarray datasets used in geWorkbench can be clustered based
on expression profiles for genes, for single arrays, or both. For example in clustering by gene, if the expression pattern for gene A across all experiments is very similar to that of gene B, then A and B will tend to cluster together.

**Example**

1. Load a microarray dataset, for example `web100.exp` as shown in Chapter 3 of this manual.

2. In the **Array/Phenotype** component at lower left in the GUI, select which arrays should be used in the analysis (Figure 4-1). Here we have activated all the arrays by checking the boxes next to each group. (The same could be accomplished by not checking any boxes, since the default is to include all arrays if nothing is selected).

   ![Figure 4-1 Activating the array sets](image_url)

3. In the **Analysis** tab at lower right in the GUI, select **Hierarchical Clustering**. Select the **Services** tab (Figure 4-2). Click on the blue text **Change Index Service**. This will bring up a small input box.
4. For **host**, (if it is not already present) enter `cagridnode.c2b2.columbia.edu` (Figure 4-3). Leave the port set to 8080. Press **OK**.

5. Pressing **Grid Services** will display the available services (Figure 4-4). Select the available **Hierarchical Clustering** service. Once a service is selected, its details will display in the **Service Details** box below.
6. Now go back to the **Parameters** tab (see Figure 4-1 above) and set the following parameters for this example clustering operation:
   a. Clustering Method: **Total Linkage**
   b. Clustering Dimension: **Both**
   c. Clustering Metric: **Pearson’s**

7. Click **Analyze**. The computation is carried out on the remote server and returned to geWorkbench, where the results are entered into the Project as a new data node (Figure 4-5), and displayed in the **Dendrogram Viewer** component (Figure 4-6).

---

**Figure 4-4** Selecting an available Hierarchical Clustering service

**Figure 4-5** Result sets displayed in the Project Folder
8. Within the Dendrogram display, the data can be manipulated in many ways, and clusters of genes can be selected and stored as named sets in the Marker component for further analysis (see the Clustering tutorial on http://www.geworkbench.org/ for details).

Self-Organizing Maps (SOM)

The SOM algorithm is used to divide a data set into a predetermined number groups based on similarity. Here we illustrate running SOM through the grid interface.

Example

1. Start as in the previous example by loading a microarray dataset, such as web100.exp. If it is already loaded, there is no need to reload it.

2. In the Analysis tab, select SOM.

3. Changing the Index Service, which should not be necessary, has been described above under Hierarchical Clustering.

4. Select an available SOM service. Once selected, the details will display in the area below it (Figure 4-7).
5. We will accept the default parameters, except setting the Function to Gaussian instead of Bubble (Figure 4-8):

   a. **Rows**: 3
   b. **Columns**: 3
   c. **Radius**: 3
   d. **Iterations**: 4000
   e. **Alpha**: 0.8
   f. **Function**: Gaussian
Figure 4-8 Setting SOM parameters

6. Click **Analyze**. The result will be returned from the remote server and displayed in the SOM Viewer component as a 3x3 array of graphs. Each contains a set of expression profiles that are, within the boundaries established by the parameters chosen, similar to each other. Several clearly different groups can be distinguished (Figure 4-9).

Figure 4-9 SOM Viewer
7. The genes represented in any given graph can be selected and returned to the Markers component for further analysis.

8. The result is also placed in the Project Folders component of the GUI beneath its parent data set. In this figure, both a hierarchical clustering dataset and a SOM result are present. Either can be viewed simply by selecting it (Figure 4-10) here.

![Figure 4-10 Project Folder with results nodes](image)

**ARACNE**

**Introduction**

ARACNE (Algorithm for the Reconstruction of Accurate Cellular Networks) can be used to infer regulatory interactions within a set of microarray data. Its use has been described in detail in reference Margolin et al. 2006, #1.

The ARACNE algorithm is based on the calculation of mutual information (MI). It has been designed to overcome a number of problems with other methods, for example by allowing calculation on continuous-valued data rather than requiring discretized data, and needing no assumptions about the underlying network topology. The method is not without potential limitations, and the user should consult the references for more information on the usefulness of the algorithm in any particular type of investigation.

ARACNE calculates the mutual information between specified pairs of markers across a set of multiple microarray gene-expression experiments. In an optional second step, an
information theoretic property known as the Data Processing Inequality (DPI) (Margolin et al. 2006, #2) is used to attempt to remove indirect interactions, that is, those mediated by another marker. The algorithm in principal could be used on any type of interaction data, not just gene expression. In geWorkbench however only gene expression data may currently be submitted to the algorithm. The output of ARACNE is an adjacency matrix, showing the strength of interaction for each calculated pair of markers.

Prerequisites:

There should be at least 100 microarrays in the dataset to allow reliable calculation of the mutual information. However, please note than on such large datasets, only calculation of the mutual information of a particular gene marker with the rest of the dataset should be undertaken on a desktop-class machine. An all-against-all calculation generally requires use of a cluster computer, potentially via the grid service facility if such a service becomes available. Otherwise, ARACNE can be obtained as a stand-alone program for use on a local computational cluster.

The data used should show a significant dynamic range, for example through sampling multiple phenotypes, or through use of experimental perturbations. Uninformative genes, such as those with low mean expression, should likely be filtered out before running ARACNE.

Understanding the Parameters

There are several choices that can be made as to how ARACNE will be run on a given dataset. Here we will outline what these choices are, and provide an introduction on how to choose appropriate parameters for your own particular case. The figure below (Figure 4-11) shows the ARACNE component within geWorkbench and the parameters which can be set.
Hub Markers – Using a hub gene to limit calculations

An ARACNE run can either compute the MI values of every (active) marker against every other, or it can use a list of one or more hub genes to limit the number of calculations required. For large datasets, the all-against-all calculation is not feasible on an local desktop machine, and instead a computational cluster should be used if available as a grid service. If hub genes are used, the MI values are only calculated between each hub gene and all others (or all other markers activated in the Markers component).

- Options:
  - All vs. All – Compute the mutual information of each marker against all others (or all markers activated in the Markers component against each other).
  - List – Compute the mutual information of each marker in the list against all others (or all those activated in the Markers component).

Threshold Type – P-value or Mutual Information Score

The threshold value above which potentially interacting pairs will be reported can be set either as a p-value or directly as a mutual information score. The mutual information score is more useful when reevaluating an already calculated adjacency matrix, an option present in the standalone version of ARACNE. Margolin et al. 2006, #1 gives a suggested method of choosing a reasonable p-value:

Divide the desired number of false-positives (generally a small integer) by the number of tests performed, calculated as the number of distinct probe pairs. For example, a
threshold of 1e-7 will lead to about five expected false-positives for a data set with around 10,000 probes, because 10,000 choose 2 (i.e., about 5e7) probe pairs are tested”.

- Options:
  - **Mutual Info** – use a specified mutual information value, ranging from 0 to 1.
  - **P-Value** – specify a p-value. This will be converted internally to a mutual information threshold as described above and in Margolin et al. 2006, #1.

**Kernel Width** – a parameter for the Mutual Information calculation

The parameter used in the mutual information calculation is the kernel width of the Gaussian operator. ARACNE will calculate a default value based on the sample size. The user may also specify a kernel width; methods to calculate an optimal value are given in Margolin et al. 2006, #1. However, for most uses the default value should be adequate, as the algorithms has been shown to be robust with respect to this parameter.

- Options:
  - **Inferred** – accept the default value
  - **Specify** – enter an explicit value.

**DPI Tolerance** – Removing indirect interactions using DPI

As described in Margolin et al. 2006, #1, “Many statistical dependencies between gene expression profiles arise from cascades of transcriptional interactions that correlate the expressions of many genes that do not interact directly. ARACNE provides an option to eliminate interactions that are likely to be indirect by applying …the DPI (described in detail in Margolin et al. 2006, #2). The DPI requires accurate estimation of Mutual Information (MI) ranks; as MI values cannot be estimated exactly with finite data, a tolerance is used to compensate for errors in the estimate that might affect these ranks. Empirically, values between 0 (no tolerance) and 0.15 (15%) tolerance should be used, as larger values tend to cause high false-positive rates”.

- Options:
  - **Do Not Apply** – do not run the DPI calculation
  - **Apply** – run the DPI calculation with the specified tolerance to remove potential indirect interactions.

**DPI Target List** (checkbox) – Generating a network of transcription factors

If you wish to reconstruct a network only involving transcription factors, you can include a list of such genes (which have been annotated as transcription factors (TFs)) whose interactions are not to be eliminated by DPI in favor of interactions.
consisting of two non-TFs. “This partially alleviates the problem associated with highly correlated non-interacting genes, such as those involved in stable complex formation, which violate some of the assumptions required for application of the DPI. This feature is described in greater detail in the online Supplementary Manual” (Margonlin et al. 2006, #1) (http://amdec-bioinfo.cu-genome.org/html/ARACNE.htm).

Running an ARACNE Calculation

Example

1. A microarray dataset must be loaded in the Projects component. Review the Prerequisites section above for details.
2. The ARACNE component can be found in the Analysis section (tab).
3. Set the parameters depending on the desired type of run as described above in “Understanding the Parameters”
4. If a remote ARACNE grid service is to be used, go to the Services tab in the ARACNE component and chose the desired service, e.g. as already described above for Hierarchical Clustering.
5. Push the Analyze button. When complete, the results will be displayed within geWorkbench in the Cytoscape component.

Viewing the Results

The figure below (Figure 4-12) shows an example of viewing the results of an ARACNE run using a single hub gene. The results are displayed in the Cytoscape component. This component has a number of options for controlling how the network graph will be displayed. It is an external component to geWorkbench and full documentation on its use can be found at http://www.cytoscape.org/.
Figure 4-12 Display of ARACNE-generated adjacency matrix in Cytoscape

References


Chapter 5  Querying caARRAY

This chapter describes how geWorkbench can query remote instances of a caARRAY database. caARRAY uses a MAGE-OM compliant API which allows searching on a number of common annotation data fields, such as species, array type and tissue type.

This chapter covers the single topic:
- Searching caARRAY using MAGE annotations

Searching caARRAY using MAGE annotations

caARRAY is a microarray gene expression repository developed by the NCICB which supports storing and querying of annotated datasets. The annotations conform to the MAGE (Microarray Gene Expression) model. It should be noted that actual datasets may be only partially or sparsely annotated, but the model has the capability to store, in a rigorous fashion, almost any type of information about an experiment.

In the current implementation, geWorkbench can query against four types of annotations supported by caARRAY:
- Tissue type (available choices are displayed)
- Chip Platform (e.g. Affymetrix, Agilent etc.)
- Organism
- Principal Investigator

The following example illustrates constructing a query against caARRAY.

1. Create a new project as described previously.
2. Right-click on the Project entry and select Open File(s).
3. Click the Remote radio button. This will cause the Open File popup to switch to the remote file interface (Figure 5-1).
You can inspect the **caARRAY Index Service** tab (Figure 5-2). As more services become available, the desired service can be entered here:

![Open File dialog](image)

*Figure 5-1 The Remote Open File interface*

Clicking on the red **Go** button would retrieve all available experiments. Instead, we will construct a query.

4. **Click on Query** (see Figure 5-1 above) to build a MAGE keyword search. The query builder appears (Figure 5-3).
5. Under Category, select Experiments (Figure 5-4). The available search field types will be displayed. Here we will search on Tissue Type. Highlighting this field shows available tissue types.

6. Here we have selected the tissue type ovary.

Notes on query building –
10. Clicking on **Get bioassays** returns the available bioassays for this experiment. (This hidden step is the most easily overlooked one in geWorkbench). The
returned bioassays are shown in Figure 5-6.

Figure 5-6 View of retrieved bioassays

11. Click on the **Merge Files** checkbox at lower left (Figure 5-6) so that the four files will be merged into a single geWorkbench dataset after they are downloaded. Highlight the desired bioassays.

12. Click **Open** (located in the **Experiment Information** box). The datasets are downloaded, merged, and inserted into the current **Project** (Figure 5-7). In the **Arrays/Phenotypes** component below, the four individual arrays can be seen.
13. The merged dataset can be renamed if desired by right-clicking on it and selecting **Rename**.

14. Did you forget to check the **Merge** checkbox before download? You can merge the files after download by selecting menu item **File > Merge Datasets**.
Chapter 6 Using caSCRIPT to automate actions

This chapter describes how caSCRIPT, a scripting language developed for geWorkbench, can be used to automate common or complex tasks in geWorkbench. The caSCRIPT environment includes a visual editor with access to the available methods and variables.

This chapter covers the single topic:

- Using caSCRIPT to automate tasks

Using caSCRIPT to automate tasks

geWorkbench has a built-in scripting language, caSCRIPT. This language is similar to Java. The actual language is described in a separate technical document. It provides direct access to all of the modules in geWorkbench. Scripting allows any sequence of steps in a workflow to be automated, and allows them to be repeated as desired. The example presented here will illustrate how caSCRIPT can be used to execute a caGRID-based SOM calculation. The basics of running a SOM calculation on a caGRID node have already been covered above in Chapter 4.

The caSCRIPT component, shown below in Figure 6-1, contains a default demonstration script. It executes SOM clustering. A line to run Hierarchical Clustering has been commented out, but can be included by simply removing the comment symbol (//). For this example the user can paste in a script which will perform hierarchical clustering:

```java
void main() {
    // Instantiate project panel and cagrid panel
    module projectWindow projectPanel;
    module expressionFileFilter expFileFormat;
    module cagrid cagrid;
    string urls[1];

    // Load a microarray set
    projectPanel.loadDataSet("data/web100.exp", expFileFormat);
    datatype DSMicroarraySet mset = projectPanel.getDataSet();

    // Get services
    string url = cagrid.getServiceUrl("cagridnode.c2b2.columbia.edu", 8080,
            "HierarchicalClustering");
    print url;
```
// Do clustering
datatype DHierClusterDataSet cluster =
cagrid.doClustering(mset, "Total", "Both", "Pearson", url);
print cluster.getLabel();

// Add cluster to project panel
projectPanel.addDataSetNode(cluster);
}

The caSCRIPT component contains three separate areas (Figure 6-1) – the script display at left, a list of available methods at top-right, and an area to display the details of any selected method at bottom right. These aid in constructing new scripts (not covered in this manual).

![Figure 6-1 The caSCRIPT interface](image)

Figure 6-1 The caSCRIPT interface

Figure 6-2 below shows an example of listing a local method, which displays its properties just below it.
Figure 6-2  caSCRIPT methods selector

The Grid tab at right on the caSCRIPT component (Figure 6-3) reveals an interface for changing the caGRID Index Service and for discovering the analytical services the chosen node offers. The figure below shows that after the Discover button has been pushed, the Hierarchical Clustering service is offered. This information could be used in constructing a new script. When a discovered service is selected, its details are displayed in the area below.

Figure 6-3  Discovering available services

The caSCRIPT component has four main controls, shown below in Figure 6-4. The are
from left:

**Play** – execute the current script.

**Stop** – stop execution of the script.

**Save** – save the current script to disk.

**Open** - opens a file browser to locate a saved script.

---

**Figure 6-4  caSCRIPT controls**

**Example:**

1. To run the hierarchical clustering example, select and copy the test script shown above into the caSCRIPT component.

2. Press the left-arrow shaped **Play** button on the caSCRIPT component. The task will be executed on the remote server and the results returned to the Dendrogram component, as shown in Figure 6-5 below.
Figure 6-5  The Dendrogram component displaying results of hierarchical clustering executed using caSCRIPT

The result of running the default script, which executes SOM clustering, is shown in Figure 6-6 below:
Figure 6-6 Results of SOM clustering example using caSCRIPT
Chapter 7  Error Messages/Indicators and Problem Resolutions

Why is the desired caGRID service not available?

Until services are installed in an official caBIG index service, their availability may vary.

After running a large local calculation, my Windows computer seems slow.

There are apparently some problems, at least with Windows XP, if the operating system is pushed into swapping – that is, some of the contents of memory are written to disk. Even after geWorkbench has been exited, the slow behavior may persist. In extreme cases, simply reboot the computer. We recommend running geWorkbench in a computer with at least 1 GB of memory, while 2 GB or more will greatly increase the size of calculations possible.

How do I increase the memory allocated to geWorkbench?

There is a file in the geWorkbench root directory called UILauncher.lax. There is a line there which specifies the Java heap size:

```
lax.nl.java.option.java.heap.size.max=640678989
```

Here it is shown set to about 640 MB. You can experiment with increasing this, subject to the amount of memory in your machine and demands on it from other applications.

(Nota – this method applies to the packaged distribution version of geWorkbench)

Where else can I look for help?

Please see the main geWorkbench website at [http://www.geworkbench.org/](http://www.geworkbench.org/). Of particular interest will be the following sections:

1. FAQs
2. Known Issues
3. Tutorials
Appendix A References

Scientific Publications


Technical Manuals/Articles


3. Extensible Markup Language: http://www.w3.org/TR/REC-xml/


caBIG Material

1. caBIG: http://cabig.nci.nih.gov/

2. caBIG Compatibility Guidelines: http://cabig.nci.nih.gov/guidelines_documentation
caCORE Material

1. caCORE: http://ncicb.nci.nih.gov/core
2. caBIO: http://ncicb.nci.nih.gov/core/caBIO
3. caDSR: http://ncicb.nci.nih.gov/core/caDSR
4. EVS: http://ncicb.nci.nih.gov/core/EVS
5. CSM: http://ncicb.nci.nih.gov/core/CSM
Appendix B Glossary

Following is a list of terms and their definitions.

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<th>Definition</th>
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<tr>
<td>API</td>
<td>Application Programming Interface</td>
</tr>
<tr>
<td>caArray</td>
<td>cancer Array Informatics</td>
</tr>
<tr>
<td>caBIG</td>
<td>cancer Biomedical Informatics Grid</td>
</tr>
<tr>
<td>caBIO</td>
<td>Cancer Bioinformatics Infrastructure Objects</td>
</tr>
<tr>
<td>caCORE</td>
<td>cancer Common Ontologic Representation Environment</td>
</tr>
<tr>
<td>caDSR</td>
<td>Cancer Data Standards Repository</td>
</tr>
<tr>
<td>caMOD</td>
<td>Cancer Models Database</td>
</tr>
<tr>
<td>CDE</td>
<td>Common Data Element</td>
</tr>
<tr>
<td>CGAP</td>
<td>Cancer Genome Anatomy Project</td>
</tr>
<tr>
<td>CMAP</td>
<td>Cancer Molecular Analysis Project</td>
</tr>
<tr>
<td>CVS</td>
<td>Concurrent Versions System</td>
</tr>
<tr>
<td>EVS</td>
<td>Enterprise Vocabulary Services</td>
</tr>
<tr>
<td>GUI</td>
<td>Graphical User Interface</td>
</tr>
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<td>HTTP</td>
<td>Hypertext Transfer Protocol</td>
</tr>
<tr>
<td>JAR</td>
<td>Java Archive</td>
</tr>
<tr>
<td>Javadoc</td>
<td>Tool for generating API documentation in HTML format from doc comments in source code (<a href="http://java.sun.com/j2se/javadoc/">http://java.sun.com/j2se/javadoc/</a>)</td>
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<tr>
<td>MAGE</td>
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<tr>
<td>MAGE-OM</td>
<td>MicroArray Gene Expression - Object Model</td>
</tr>
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<td>MGED</td>
<td>Microarray Gene Expression Data</td>
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<tr>
<td>MO</td>
<td>MGED Ontology</td>
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<tr>
<td>NCI</td>
<td>National Cancer Institute</td>
</tr>
<tr>
<td>NCICB</td>
<td>National Cancer Institute Center for Bioinformatics</td>
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<td>SDK</td>
<td>Software Development Kit</td>
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<td>SQL</td>
<td>Structured Query Language</td>
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